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PHYSICOCHEMICAL ANALYSIS OF SUNFLOWER HONEY FROM BULGARIA

Bulgaristan Ayçiçek Ballarının Fiziko-Kimyasal Analizi

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ABSTRACT

Physicochemical properties of 27 sunflower honey samples from Bulgaria were investigated. The botanical origin of the samples was ascertained by pollen analysis. The honey samples displayed relative frequencies of *Helianthus annuus* L. pollen up to 41%. The ranges for water content (15.60–19.30%), reducing sugars (72.51–80.80%), sucrose (0.50–3.70%), diastase (9.00–20.80 Gothe units), hydroxymethylfurfural (HMF), (0.69–12.40 mg/kg), total acidity (17.70–36.00 meq/kg), electrical conductivity (0.23–0.48 mS/cm), proline (218.50 – 679.50 mg/kg), specific rotation (-20.20–(-12.30)) $[\alpha]_D^{20}$ were obtained. The results obtained also suggest that these honey samples are of good quality. The results are in agreement with standards of quality established by national and international regulations. Significant moderate correlation between electrical conductivity and specific rotation was found ($r=0.582$, $p<0.05$).

Keywords: Honey, sunflower honey, physicochemical properties, quality parameters

ÖZ

Bulgaristan'dan alınan 27 ayçiçeği balı örneğinin fiziko-kimyasal özellikleri araştırıldı. Örneklerin botanik kökeni polen analizi ile belirlendi. Bal numuneleri, %41'e varan oranlarda *Helianthus annuus* L. polen sıklığı göstermiştir. Su içeriği (%15.60 – 19.30), indirgeyici şekerler (%72.51 – 80.80), sakaroz (%0.50 – 3.70), diastaz (9.00 – 20.80 Gothe birimleri), hidrosimetilfurfural (HMF), (0.69 – 12.40mg/kg) için aralıklar, toplam asitlik (17.70 – 36.00meq/kg), elektriksel iletkenlik (0.23 – 0.48 mS/cm), prolin (218.50 – 679.50 mg/kg), spesifik rotasyon (-20.20 – (-12.30)) $[\alpha]_D^{20}$ elde edildi. Elde edilen sonuçlar da bu bal örneklerinin kaliteli olduğunu göstermektedir. Sonuçlar, ulusal ve uluslararası düzenlemeler tarafından belirlenen kalite standartları ile uyumludur. Elektriksel iletkenlik ile özgül rotasyon arasında orta düzeyde anlamlı korelasyon bulundu ($r=0,582$, $p<0,05$).

Anahtar Kelimeler: Bal, ayçiçeği balı, fiziko-kimyasal özellikler, kalite parametreleri

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GENİŞLETİLMİŞ ÖZET

Giriş: Çiçek balı, bal arıları (*Apis mellifera* L.) tarafından bitkilerin nektarından üretilen doğal bir maddedir. Bal, tek çiçekli bitki veya çok bitki kaynağı olarak sınıflandırılır. Tek çiçekli bitki balı, ağırlıklı olarak nektar ve polen içeren bir bitki türü tarafından üretilir. Farklı çiçekli bitki balları, tipik melissopalınolojik ve fizikokimyasal özellikler gösterir. Çok kaynaklı bitki balı, hiçbir baskın olmayan çeşitli bitkilerden nektar ve polenlerden üretilir. Bu çalışma, Bulgaristan'da üretilen ayçiçeği balının fizikokimyasal parametrelerini değerlendirmeyi amaçlamaktadır. Bulgaristan'dan alınan 27 ayçiçeği balı örneğinin fizikokimyasal özellikleri araştırıldı.

Gereç ve Yöntem: Bal örnekleri oda sıcaklığında cam kaplarda muhafaza edilmiştir. Çalışmada 2017 ve 2018 arıcılık sezonlarında hasat edilen arı balının temsili örnekleri kullanılmıştır. Örneklerin botanik kökeni polen analizi ile tespit edilmiştir. Bal numuneleri, %41'e varan oranlarda *Helianthus annuus* L. polen sıklığı göstermiştir. Bulgaristan Arı Balı Devlet Standardı 2673-80'e göre *H. annuus*'tan %40'a kadar polen içeren bal örnekleri ayçiçeği balı olarak sınıflandırılabilir. Analizler, numunelerin laboratuvara teslim edildiği tarihten itibaren bir ay içinde yapılmıştır. Polen analizi ve fizikokimyasal analizler Bulgaristan, Sofya'daki Veteriner Kontrol ve Ekoloji Merkez Laboratuvarında yapıldı.

Bulgular: Ayçiçeği balının şu parametreleri Avrupa Bal Komisyonu tarafından önerilen yöntemlere göre belirlendi: su içeriği, indirgeyici şekerler, sakaroz, diastaz, HMF, toplam asitlik, elektriksel iletkenlik, prolin ve spesifik rotasyon. Su içeriği (%15.60 – 19.30), indirgeyici şekerler (%72.51 – 80.80), sakaroz (%0.50 – 3.70), diastaz (9.00 – 20.80 Gothe birimleri), hidrosimetilfurfural (HMF), (0.69 – 12.40mg/kg) için aralıklar), toplam asitlik (17.70 – 36.00meq/kg), elektriksel iletkenlik (0.23 – 0.48 mS/cm), prolin (218.50 – 679.50 mg/kg), spesifik rotasyon (-20.20 – (-12.30)) elde edildi. Spesifik optik rotasyon, bal örneklerinin çiçek balından geldiğini, dönen negatif olduğunu (laevorotatory) gösterir. Numunelerin çoğu 350 mg/kg'a kadar prolin değerlerine sahiptir. HMF ortalama 2.82 mg/kg olarak belirlendi. Elde edilen sonuçlar da bu bal örneklerinin kaliteli olduğunu göstermektedir. Sonuçlar, ulusal ve uluslararası düzenlemeler tarafından belirlenen kalite standartları ile uyumludur. Elektriksel iletkenlik ile özgül rotasyon

arasında orta düzeyde anlamlı korelasyon bulundu ($r=0,582$, $p<0,05$).

Sonuç: Sonuç olarak, tüm bal örnekleri %40'tan fazla *H. annuus* polen içerir ve ayçiçeği balı olarak sınıflandırılabilir. Ayçiçeği balının su içeriği, indirgeyici şekerler, sakaroz, diastaz, HMF, toplam asitlik, elektriksel iletkenlik, prolin ve özgül rotasyon parametreleri uluslararası yönetmeliklerle belirlenen kalite parametrelerini karşılamaktadır. Bu çalışma, Bulgaristan'dan ayçiçeği balının karakterizasyonu için faydalı bilgiler sağlamaktadır.

INTRODUCTION

Blossom honey is a natural substance produced by honey bees (*Apis mellifera* L.) from the nectar of plants. Honey is classified as unifloral or polyfloral. The unifloral honey is produced by one plant species containing predominantly its nectar and pollen. The different unifloral honeys show typical melissopalynological and physicochemical properties. The polyfloral honey is produced from nectar and pollen from various plants, none of which is predominant. In general, unifloral honeys are regarded as more valuable products. They have good quality and specific sensorial characteristics. Furthermore, market prices of honey are determined by its botanical origin. It can be said that monofloral honeys are more expensive than mixed polyfloral honeys. This is due to their pharmacological and organoleptic properties (Silva et al. 2009, Bilandžić et al. 2017). Thus, great attention has been paid for identification of unifloral honey by researchers (Wen et al. 2017). Honey is becoming an effective therapeutic agent by practitioners of conventional medicine due to its chemical composition. In this respect the composition of honey depends on its floral origin (De-Melo et al. 2018, Valdés-Silverio et al. 2018). According to Ahmed et al. (2016) and Saxena et al. (2010) honey includes a lot of substances. The main constituents of honey are carbohydrates and water. Phenolic compounds, amino acids, vitamins, minerals and enzymes are also presented in honey.

In Bulgaria there are low levels of local consumption of bee honey – under 200 g on average per capita compared to some countries in Western Europe (about 1 kg on average per year). Bulgarian beekeeping has a traditionally export character. According to Agro-statistics of the Ministry of agriculture and food the export of Bulgarian honey

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for the period 2015 – 2017 increased from 9784 to 13302 tons per year (Aleksiev 2019). In Bulgaria very often the sunflower honey is mixed with polyfloral honey and it is sold as polyfloral honey.

Different unifloral honey types are produced in Europe. In Bulgaria, one of the most important honey types are black locust honey (*Robinia pseudoacacia*), lime honey (*Tilia spp.*), rape honey (*Brassica spp.*), sunflower honey (*H. annuus*). Some honey types such as black locust, lime, rape coriander, fennel honey were studied and described by Atanassova et al. (2012), Dinkov (2014) while data for others such as sunflower honey is insufficient. Sunflower is largely cultivated in many European countries because it represents to bees nectar and pollen (Persano Oddo and Piro 2004). To date, several studies have attempted to elucidate the sunflower honey produced in Bulgaria (Nikolova et al. 2014, Atanassova et al. 2012). The present study aims to evaluate physicochemical parameters of sunflower honey produced in Bulgaria. The obtained results were compared with national (Bulgarian State Standard for Bee Honey 3050-80 and Bulgarian State Standard 2673-80) and international regulations (Council Directive 2001/110 relating to honey (2002) and Codex Alimentarius (2001)).

MATERIALS AND METHODS

In this study, 27 sunflower honey samples were obtained mainly from Northwest, North and West Bulgaria. The honey samples (about 500 g) were kept in glass containers at room temperature. The study used representative samples of bee honey harvested in beekeeping seasons 2017 and 2018. The analyses were performed within one month from the date of receiving the samples at the laboratory. The pollen analysis was carried out by Bulgarian State Standard for Bee Honey 3050-80 and Bulgarian State Standard 2673-80. According to Bulgarian State Standard for Bee Honey 2673-80 honey samples with more than 40% *H. annuus* pollen is sunflower honey. The pollen analysis and physicochemical analyzes were done at the Central Laboratory of Veterinary Control and Ecology, Sofia, Bulgaria.

The physicochemical parameters water content, diastase, reducing sugars and sucrose, hydroxymethylfurfural, total acidity and electrical

conductivity, proline content and specific rotation were determined according to the European Honey Commission recommended methods (Bogdanov et al. 1997):

-Water content is determined by the refractometric method (Abbe refractometer);

-Determination of diastase activity is after Schade method;

-For reducing sugars and sucrose is used Fehling's reagent;

-Hydroxymethylfurfural was determined after White (in mg/kg);

-Total acidity (meq/kg) by titration with 0.1 N sodium hydroxide with phenolphthalein indicator;

-Electrical conductivity was measured in 20% weight volume in water (the results are expressed in mS/cm);

-Proline content was determined spectrophotometrically. Proline stock solution with 0.8 mg/mL concentration was prepared.

Statistical analysis of the results was performed using SPSS 20.0 for Windows. Correlations between results were made using the Pearson's correlation coefficient (r), ($p < 0.05$). All results are presented as minimal and maximal value, means \pm standard deviation.

RESULTS

Melissoplainological analysis is based on the identification of pollen by microscopic examination, and it needs highly specialized personnel. Another limitation of the method is that sometimes the pollen grains have similar morphologies and it is difficult to be recognized (Bogdanov et al. 2004). In many cases the melissoplainological analysis presents the dominant pollen type in the honey. In general, all honey samples could be considered as sunflower honey according to their pollen content (up to 41% *H. annuus* pollen). According to Bulgarian State Standard for Bee Honey 2673-80 honey samples with up to 40% pollen samples from *H. annuus* can be classified as sunflower honey. The main species are presented in Table 1. The *H. annuus* pollen varies in the ranges 41 – 80%.

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Table 1. Pollen analysis of studied honey samples (n=27)

Honey samples	Main species or family, %±SD
1	Asteraceae 80.1±4.2 from them <i>Helianthus annuus</i> – 72.1±3.4
2	Asteraceae 40.4±2.1 from them <i>Helianthus annuus</i> – 43.3±1.7
3	Asteraceae 84.5±4.4 from them <i>Helianthus annuus</i> – 71.8±4.3
4	Asteraceae 52.8±1.2 from them <i>Helianthus annuus</i> – 41.7±0.5
5	Asteraceae 63.8±3.3 from them- <i>Helianthus annuus</i> – 60.7±3.2
6	Asteraceae 70.3±3.7 from them- <i>Helianthus annuus</i> – 69.0±3.6
7	Asteraceae 52.9±2.8 from them <i>Helianthus annuus</i> – 48.7±2.5
8	Asteraceae 57.1±3.0 from them <i>Helianthus annuus</i> – 49.3±2.6
9	Asteraceae 74.9±3.9 from them <i>Helianthus annuus</i> – 73.9±3.8
10	Asteraceae 49.5±2.6 from them <i>Helianthus annuus</i> – 48.4±2.5
11	Asteraceae (<i>Helianthus annuus</i> – 69.3±4.6); Apiaceae (<i>Coryandrum sativum</i> – 3.8±0.2); Brassicaceae –2.5±0.2
12	Asteraceae (<i>Helianthus annuus</i> – 52.1±4.3); Asteraceae (<i>Cardus nutans</i> – 8.0±0.4)
13	Asteraceae (<i>Helianthus annuus</i> – 64.7±3.4); Brassicaceae – 9.0±0.5; Apiaceae (<i>Coryandrum sativum</i> –7.7±0.4)
14	Asteraceae (<i>Helianthus annuus</i> – 62.1±1.3); Apiaceae (<i>Coryandrum sativum</i> – 4.5±0.2); Asteraceae (<i>Cardus nutans</i> – 4.2±0.2)
15	Asteraceae (<i>Helianthus annuus</i> – 62.1±3.3)
16	Asteraceae (<i>Helianthus annuus</i> – 49.5±2.5)
17	Asteraceae (<i>Helianthus annuus</i> – 63.9±1.3)
18	Asteraceae (<i>Helianthus annuus</i> – 50.0±2.5)
19	Asteraceae (<i>Helianthus annuus</i> – 74.1±1.7)
20	Asteraceae (<i>Helianthus annuus</i> – 52.5±1.8)
21	Asteraceae (<i>Helianthus annuus</i> – 57.1±2.3)
22	Asteraceae (<i>Helianthus annuus</i> – 70.2±4.1)
23	Asteraceae (<i>Helianthus annuus</i> – 66.2±2.0)
24	Asteraceae (<i>Helianthus annuus</i> – 71.5±3.6)
25	Asteraceae (<i>Helianthus annuus</i> – 55.4±1.7)
26	Asteraceae (<i>Helianthus annuus</i> – 57.9±2.0)
27	Asteraceae (<i>Helianthus annuus</i> – 72.7±1.4)

Table 2 presents the results of the analysis of the honey samples: the average values and standard deviation of the physicochemical parameters (water

content, reducing sugars, sucrose, diastase, HMF, total acidity, electrical conductivity).

Table 2. Physicochemical parameters of sunflower honey, n=27

Parameters	Min	Max	Mean±SD
Water content, %	15.60	19.30	17.76±0.96
Reducing sugars, %	72.50	80.80	76.15±2.60
Sucrose, %	0.50	3.70	1.72±1.01
Diastase, Gothe units	9.00	20.80	14.50±3.94
Hydroxymethylfurfural (HMF), mg/kg	0.69	12.40	2.82±2.65
Total acidity, meq/kg	17.70	36.00	23.73±7.23
Electrical conductivity, mS/cm	0.23	0.48	0.32±0.07

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Taking into consideration that the honey water content has to be lower than 20% in general (Council Directive 2001/110 relating to honey, 2002 and Codex Alimentarius, 2001), the values obtained in this study were satisfactory. They ranged from 15.60 to 19.30%.

The main reducing sugars in honey are glucose and fructose. The results for reducing sugars were considered to be sufficient for identification of honey quality. The mean value and standard deviation for the reducing sugars and sucrose are determined as $76.15 \pm 2.60\%$ and $1.72 \pm 1.01\%$, respectively. The results for the minimal and maximal values are presented in Table 2.

According to international regulation (Council Directive 2001/110 relating to honey (2002)) diastase activity must not be less than 8 Gothe units. All results are up to this value. In the present study the diastase activity varied by a large range (Table 2).

Hydroxymethylfurfural (HMF) was determined as 2.82 mg/kg on average. The highest value is under 13 mg/kg.

The mean total acidity value was below 50 meq/kg of honey and satisfied the European regulation for this parameter (Council Directive 2001/110 relating to honey (2002)). The total acidity ranged from 17.70 to 36.00 meq/kg.

As expected, sunflower honey had the lower values of electrical conductivity (average 0.32 mS/cm). Significant moderate correlation between electrical conductivity and specific rotation was found ($r=0.582$, $p<0.05$).

The measured proline content of sunflower honey is shown on Figure 1. In the present study the proline content varies in very large ranges 218.50 – 679.50 mg/kg. The average values and standard deviation are 404.94 ± 144.78 mg/kg. As can be seen from Figure 1 most of the samples have proline values up to 350 mg/kg.

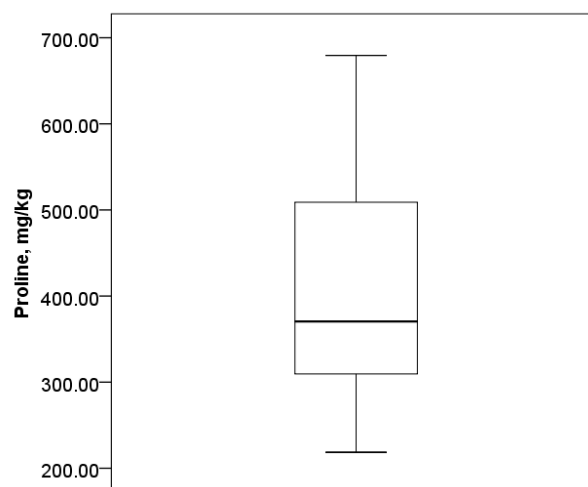


Figure 1. Box plot diagram of proline content in sunflower honey samples. Minimal, maximal and median values are presented.

Specific rotation of sunflower honey samples ranged from -20.20 to -12.30 $[\alpha]_D^{20}$ and mean value \pm standard deviation (-17.23 ± 2.43 $[\alpha]_D^{20}$). Sample 14 (value 22.20) is an outlier. The results are shown on Figure 2.

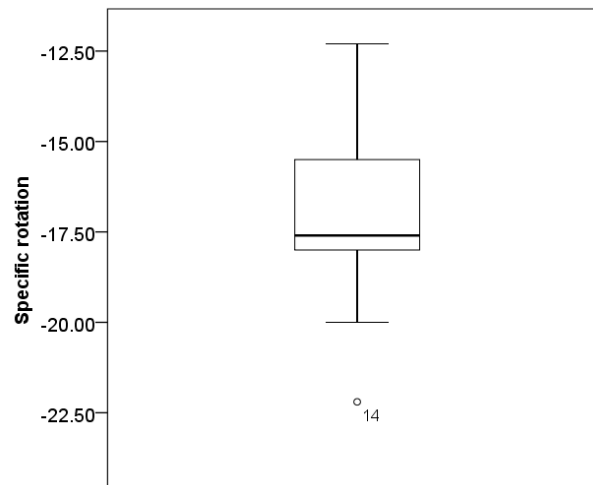


Figure 2. Box plot diagram of specific rotation on sunflower honey samples. Minimal, maximal and median values are presented

DISCUSSION

Melissopalynological analysis is very important tool in the analysis of honey. Pollen analysis is generally used to determine and confirm the botanical origin of the honey samples. The *H. annuus* pollen was

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predominant in all honey samples, accounting for more than 40% pollen grains. Oroian and Ropciuc (2017) reported that their samples had around 60% of *H. annuus* pollen. In a recent study, Pauliuc and Oroian (2020) presented more than 45% pollen grains of *H. annuus* (ranges 50 – 92% pollen grains).

Water is quantitatively one of the most important components of honey. This parameter describes honey quality. Honeys with high levels of water (more than 20% in general) tend to ferment more easily. Its content can affect the storage of honey. Furthermore, the water content in honey depends on beekeeping practices and environmental conditions. Also, it can vary from year to year. The average value ($17.76 \pm 0.96\%$) is in accordance with the results of Isopescu et al. (2014) for sunflower honey samples.

The sugar content mainly depends on nectar of the flowers used by the bees. Therefore, it varies in the different honey type. In this respect, the content of some sugars and the ratios between them are used to determine honey authenticity (Borrás et al. 2014). Some geographical and climatic conditions can also affect sugar composition of honey (Mărghitaş et al. 2009, Kaskonienė et al. 2010). Reducing sugars are the most abundant sugars in floral honey samples. The sucrose content is under the limits presented by Council Directive 2001/110 relating to honey (2002) and Codex Alimentarius (2001). Gropoşilă-Constantinescu et al. (2020) presented 74.8% reducing sugars for commercial sunflower honey samples. This result is very similar to the average value in the present study. Sahinler et al. (2009) reported higher values for sucrose content in sunflower honey ($6.46 \pm 0.78\%$) which is higher than the maximal value for the Bulgarian sunflower honey samples (3.70%).

Enzyme diastase is derived from the glandular secretions of the honey bees. Diastase activity is one of the most important quality parameter. Very often it is used to determine if honey has been heated during storage. Diastase activity loss occurs as temperature increases. The average diastase activity in this study is comparable to those presented by Gropoşilă-Constantinescu et al. (2020) for sunflower honey (about 12 Gothe units). Juan-Borrás et al. (2014) evaluated the influence of the country origin on some physicochemical parameters including diastase activity. Their results for diastase activity in sunflower honey samples from Spain, Romania and Czech Republic are very similar to the

results in the present study. The diastase activity varies not only according to its botanical origin and country origin but also due to high temperature and storage. Eremia et al. (2019) reported large ranges for diastase activity (15.02 – 30.69 Gothe units).

Hydroxymethylfurfural (HMF) can be found in low concentrations in honey. It is produced from fructose in the presence of free acids. The production of HMF depends on the temperature (Da Silva et al. 2016). Thermal treatment of honey can generate toxic HMF, thereby resulting in quality reduction. For the sunflower honey is typical quick crystallization (Persano Oddo and Piro 2004). This is due to high glucose content in this honey type. Glucose may crystallize at room temperature. For this reason, sometimes the sunflower honey is liquefied at high temperature. This over heating produces HMF. Based on the Codex Alimentarius (2001) and Council Directive 2001/110 relating to honey (2002), HMF level should not exceed 40 mg/kg. The obtained results for HMF are close to those previously reported by Sakača et al. (2019) and Pauliuc and Oroian (2020). The average HMF value and standard deviation (1.19 ± 1.12 Gothe units) reported by Sakača et al. (2019) are consistent with our results. Juan-Borrás et al. (2015) found higher HMF values (37.4 mg/kg, 37.9 mg/kg, and 39.8 mg/kg) in sunflower honey. The authors noted that these values are unacceptable for raw honey. These outlying values are not frequent.

Acidity of honey depends on the presence of organic acids and inorganic ions. Acid measurement in honey evaluates honey fermentation (Belay et al. 2013). Lower value of acid indicates absence of fermentations. The average value of total acidity is comparable to this presented by Kivrak et al. (2017) for sunflower honey.

Values for electrical conductivity higher than 0.80 mS/cm are not typical for nectar honeys according to Codex Alimentarius (2001). The obtained minimal and maximal values are in agreement with the result presented by Persano Oddo and Piro (2004). The correlation between electrical conductivity and specific rotation was also supported by Belay et al. (2013) and Pridal and Vorlova (2002).

In the present study most of the samples have proline values up to 350 mg/kg. These results are confirmed by Wen et al. (2017). They received very similar data. For example, the range of proline content in sunflower honeys is from 214.06 to 601.11 mg/kg. Czipa et al. (2012) found higher values for

proline in sunflower honey. Large proline variation for sunflower honeys is typical. Although proline content has been considered as a useful parameter of honey quality, other parameters can also be used for honey quality identification. International regulations of honey quality do not present information for the proline content in honey. The proline content varied in the different honey types (Cotte et al. 2004, Keckes et al. 2013, Wen et al. 2017).

Specific optical rotation is a parameter which may distinguish blossom honeys (negative values) and honeydew honeys (positive values), (Persano Oddo and Piro 2004). The concentration of various sugars are responsible for specific rotation of honey. The results obtained indicate that honey samples were from blossom honey, rotating negative (laevorotatory). Significant moderate correlation between electrical conductivity and specific rotation was found ($r=0.582$, $p<0.05$).

Conclusion

All honey samples have more than 40% *H. annuus* pollen and can be classified as sunflower honey. The parameters water content, reducing sugars, sucrose, diastase, HMF, total acidity, electrical conductivity, proline and specific rotation of sunflower honey satisfied the quality parameters established by international regulations. This study provides useful information for characterization of sunflower honey from Bulgaria.

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Author contribution: Vanya Manolova, Ivayla Parvina, Todorka Yankovska – Stefanova, Ralitsa Balkanska collected the honey samples. Vanya Manolova and Ralitsa Balkanska designed the study and carried out the experiments. Ralitsa Balkanska wrote the manuscript.

Ethical issue: Not applicable.

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