

CASE REPORT

J Res Vet Med. 2021: 40 (1) 73-76 DOI:10.30782/jrvm.885959

The Case of Gastrointestinal Nematodiosis Forming a Nodule in a Gazelle's (*Gazella subgutturosa*) Abomasum

Rahsan Yilmaz^{1*}

1Department of Pathology, Faculty of Veterinary Medicine, Harran University, 63200, Eyyubiye, Sanlıurfa, Turkey

Received 24-02-2021 Accepted 05-05-2021

Abstract

This study has reported the case of abomasal teladorsagiosis in gazelle (Gazella subgutturosa) that was found dead while under care at a wildlife reserve area in Sanliurfa, Turkey. Necropsy was performed on six-year-old, male gazelle, which was found dead in its natural environment and brought to the Department of Pathology, Faculty of Veterinary Medicine, Harran University. At necropsy were observed became whitish in color and thickened the plicae spirals on the abomasum mucosa. In the microscopic examination, parasites were encountered on the mucosa abomasum and lumen of the glands in lamina propria. Also in the lumen of the glands with parasited were detected necrotic debris and neutrophil leukocyte accumulation and to increase in the connective tissue between the glands. In the scannig electron microscopic examination, parasites were observed on the abomasum mucosa. This study is the first report case describing gastrointestinal nematodiosis forming a nodule in the abomasum of in a gazelle in Turkey.

Keywords: Gastrointestinal nematodiosis Gazella subgutturosa, histopathology, scanning electron microscopy, abomasitis

Introduction

Gazelle (Gazella subgutturosa) is the most common species among the Asian antelope species. It has a bigger and heavier structure than other Asian gazelles. ⁸ Today their number is assumed to be about 120,000-140,000; however, it has been classified precisely by International Union for Conservation of Nature (IUCN).^{7,12}

The genuses Ostertagia, Teladorsagia and Marshallagia are included in the family Trichostrongyloidae, which settle in the abomasum of domestic and wild small ruminants.²⁸ These parasites has a worldwide distribution, occurring in both tropical and subtropical areas of the world, and they are important cause of economic loss.^{14, 22}

The Teladorsagia spp. is generally a parasite that settles on the abomasum of domestic small ruminants such as sheep and goats. After the infectious larvae of the parasite are taken orally from nature, they stay until the last exsheaths by entering the glands in the abomasumal mucosa of the host. Hyperplastic gastritis-like clinical findings are observed in animals infected with Teladorsagia spp.²⁵ Pathological changes in the abomasum mucosa due to parasite are observed in three successive stages. These phases are in the form of tissue, lumen and healing phase.¹⁸ In the tissue phase; the infective period larvae taken by the host enter the lumen of the abomasum glands, forming macroscopic nodules with a diameter of about 2 mm. In parallel with the inflammatory reaction occurring in the abomasum, there is a decrease in the number of parietal and chief cells in the mucosa and an increase in the number of mucus producing cells. This increase in mucus cells, initially seen as focal, is observed in the mucosa as small, protruding and pale nodules. The middle part of these shaped nodules collapses in the appearance of the navel. This structure in the abomasum is considered an indication of hyperplasia bordered by close glands. At the end of this stage, due to

* Corresponding author: : Turkey Phone number: (+90) 414 318 3926 E-mail address: rahsany@harran.edu.tr

the separation of parasites from the gastric glands, healing of the damaged parts of the abomasum mucosa begins, and adult parasites in the lumen are automatically removed from the body.^{18, 26}

The larvae hatch from the eggs thrown out with the feces of infected animals with Ostertagia species 2 exsheath in the external environment. In infective L3 form matures in the abomasum and intestines of the last hosts.¹¹ The L3 larvae enter of the abomasum glands and exsheath ². While some L4 larvae wait in the hypobiotic stage in the abomasum glands, L5 larvae exit to the mucosa to mature.²⁶

The life cycle of Marshallagia is direct and except that L2 can hatch from eggs. Following ingestion, larvae burrow into the abomasal mucosa and forms gray-whitish nodules of 2-4 mm in size. The young L5 emerge from the nodules around 16 days post-infection, and egg-laying is apparent soon after.²²

The multinodular lesions of the abomasum are common macroscopic findings of these parasites. Therefore, is important problem to determine the actual parasite responsible for lesions.²⁸ The ante mortem diagnosis of parasite infections in livestock has been based on the detection of nematode eggs or larvae in the faeces by microscopic examination using the methods of flotation and/or larval culture.¹⁷ Besides for diagnosis PCR, histopathological and electron microscopic examinations are performed.^{3, 24, 25}

Case History

In this study the gazelle (Gazella subgutturosa) were brought from the Kızılkuyu Wildlife Reserve, located in Sanliurfa province in southeastern Turkey. The abomasum tissues of a gazelle six-year-old, male (Gazella subgutturosa), which was found dead in the natural habitat and submitted to the Department of Pathology, Faculty of Veterinary Medicine, Harran University for necropsy, constituted the material of the study. The abomasum tissue was fixed in 10% neutral buffered formalin for histopathological examination. The samples were dehydrated in graded ethanol and embedded in paraffin wax. The sections were cut at 5 µm thickness, mounted on glass slides and routinely stained with hematoxylin and eosin (H&E). The stained sections were examined by light microscope and the histopathological structure of the parasite and its lesion was determined. Besides scanning electron microscopy (SEM) analyses was applied to the affected abomasum tissue. The tissues were washed twice with phosphate-buffered saline (0.1 M, pH 7.4) and fixed in 2.5% glutaraldehyde for 48 h. Then the samples were treated with 1% osmium tetroxide for 1 h and dehydrated through a series of increasing concentrations of acetone (30%, 50%, 75%, and 100%, three repetitions each) and dried in a critical point drier. The

samples were coated with gold-palladium using a Polaron SC7620 sputter coater and examined with SEM (Leica, LEO 440, UK) at different magnifications.

In the necropsy, the mucosa of the abomasum was thicker than normal and covered with a transparent colored mucous exudate. In addition, on the abomasum mucosa was observed the whitish color, rough surface, nodular blisters or laminar thickening (Fig. 1).

In the microscopical examination, was observed that the mucosa was covered with desquamated necrotic, degenerate cell debris and occasional adult parasites (Fig. 2 A).Besides, parasites in different developmental stages were found in the lumen of some glands in lamina propria. The necrotic desquamated cell debris and neutrophil leukocyte deposits were also seen in gland lumens with parasites (Fig. 2 B and C). The increase in connective tissue attracted attention between the glands in lamina propria (Fig. 2 D). In the scanning electron microscopic examination, adult parasites with long cylindrical shape were observed on the abomasum surface and embedded in the mucosa (Fig. 3 A, B).

Unfortunately, the morphological-genetic examination could not be done because the presence of parasites was not observed macroscopically, but was detected in microscopic examination. In addition, examination for parasites in feces could not be performed as a long time has passed since of the animal death.

Discussion

As a result of the macroscopic, histopathological and scanning electron microscopic examinations, the findings were coherented with Ostertagia, Marshallagia and Teladorsagia causing abomasitis. Although it is not possible to say exactly which is the agent, the author thinks it is teladorsagia. Because, Teladorsagia is the predominant species in temperate regions and is a parasite with a high transmission rate during abundant rainy seasons.²¹ These parasites are a highly fecund parasite with even moderately infected sheep having very high faecal egg counts.²⁷ However, Teladorsagia spp. is considerably less fecund than these parasites and does not appear to be significantly more fecund than other nematode parasites found in temperate regions although defmitive data is lacking.^{2,23}

There are different published records of Teladorsagia spp. infection in sheep and goat.^{4, 9, 13, 16} In this case, histopathological lesions that findings such as necrotic debris on the abomasum mucosa and the presence of the parasite, inflammatory cell infiltration in the lamina propria, increase in connective tissue, parasites at different stages of development in the gland lumen, hyperplasia in the gland epithelium, were related with the parasite.⁶

Figure 1. Macroscopical appearance the abomasum. Nodule (arrow), laminar thickening (ahead arrow) on mucosa of the abomasum.

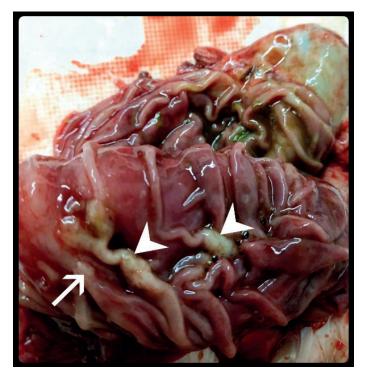


Figure 2. Microscopical appearance the abomasum:

A: Adult parasite (arrows), necrotic desquamated debris (ahead arrows), lumen (L). B and C: Parasites in different developmental stages (arrows), necrotic cells debris (N). D: Parasites in different developmental stages (arrow), increased connective tissue (CT). Hematoxylin&Eosin staining.

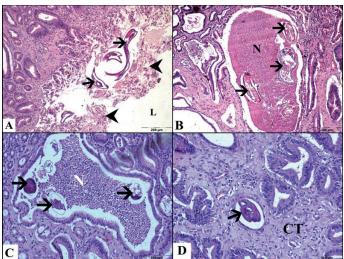
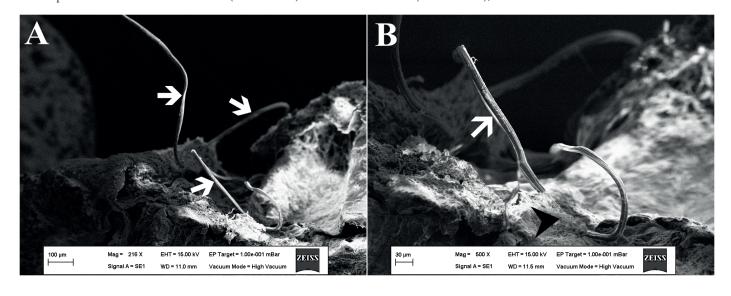


Figure 3. Scanning electron micrograpical appearance of parasite on the abomasum: A: The parasites on the abomasum mucosa (white arrows). B: Abomasum mucosa (ahead arrows),



Baker and Gershwin were stated that the cause of the inflammatory response in different layers of the abomasum could be due to the released of mediators.¹ In addition, some studies emphasized that Teladorsagia spp. increases pepsinogen release.^{1, 15} The hyperplastic changes in the glands are associated with chronic irritation during the development of the parasite in tissue¹⁹ and chemicals such as host growth factor-alpha with cytotoxic / inhibitory effect for cells and released by parasites.²⁰

Ethics

This study was carried out with permission from the Republic of Turkey Ministry of Agriculture and Forestry General Directorate of Nature Conservation and National Parks (21264211-288.04-E.1891683). This study was conducted with the approval of the Local Ethics Committee of Animal Experiments at Harran University (28.05.2020/01-19).

Conflict of Interest: No conflict of interest was declared by the authors.

References

- Baker DG, Gershwin LJ. Abomasal interstitial fluif to blood concentration gradient of pepsinogen in calves with Type-I and Type-II ostertagiosis. Am J Vet Res. 1993; 54:1294-1298.
- 2. Boag B, Thomas RJ. Epidemiological studies on gastro-intestinal nematode parasites of sheep: the seasonal nuber of generations and succession of species. Res Vet Sci. 1977; 22:62-67.
- Borji H, Raji AR, Naghibi A. The comparative morphology of Marshallagia marshalli and Ostertagia occidentalis (Nematoda: Strongylida, Trichostrongylidae) by scanning electron microscopy. Parasitol Res. 2011; 108:1391-1395.
- Cortés A, Wills J, Su X, et al. Infection with the sheep gastrointestinal nematode Teladorsagia circumcincta increases luminal pathobionts. Microbiome. 2020; 8:60.
- Dimander SO. Epidemiology and control of gastrointestinal nematodes in firstseason grazing cattle in sweden. doctoral thesis. Doctoral thesis. Swedish University of Agricultural Sciences Uppsala 2003.
- Farshid AA, Naem S, Alipour RB. Pathological changes of abomasum in naturally infected Makoyee Sheep with Teladorsagia circumcincta. Pak J Biol Sci. 2006; 9:2145-2148.
- Gurler S, Bozkaya F, Ozut D, Durmus M. Some morphological characteristics and neonatal weights of reintroduced gazelle (Gazella subgutturosa) in Turkey. Turk J Zool. 2015; 39:458-466.
- Hammond RL, Macasero W, Flores B, Mohammed OB, Wacher T, Bruford MW. Phylogenetic reanalisis of the Saudi gazelle and its implications for conservation. Conserv Biol. 2001; 15:1123-1133.
- Hoberg EP, Monsen KJ, Kutz S, Blouin MS. Structure, biodiversity, and historical biogeography of nematode faunas in holarctic ruminants: Morphological and molecular diagnoses for Teladorsagia Boreoarcticus N. sp. (Nematoda: ostertagiinae), adimorphic cryptic species in muskoxen (Ovibos Moschatus). J Parasitol. 1999; 85:910-934.
- Lichtenfels JR, Pilitt PA, Lancaster MB. Systematics of the nematodes that cause ostertagiasis in cattle, sheep and goats in North America. Vet Parasitol. 1988; 27:3-12.
- 11. Lichtenfels JR, Hoberg EP. The systematics of nematodes that cause ostertagiasis in domestic and wild ruminants in North America: an update and a key to species. Vet Parasitol. 1993; 46:33-53.
- Mallon DP. Gazella subgutturosa. The IUCN Red List of Threatened Species 2008: e.T8976A12945246. https://dx.doi.org/10.2305/IUCN.UK.2008.RLTS. T8976A12945246.en. Downloaded on 03 July 2020.

- 13. Mcneilly TN, Devaney E, Matthews JB. Teladorsagia circumcincta in the sheep abomasum: defining the role of dendritic cells in T cell regulation and protective immunity. Parasite Immunol. 2009; 31:347-56.
- 14. Molento MB. Parasite control in the age of drug resistance and changing agricultural practices. Vet Parasitol 2009;163:229-234.
- 15. Mostofa M, Mckellar QA, Eckersall PD, Gray D. Pepsinogen types in worm-free sheep and in sheep infected with Ostertagia circumcincta and Haemonchus contortus. Res Vet Sci. 1990; 48:108-111.
- Ortiz J, Ruiz de Ybáñez MR, Garijo MM et al. Abomasal and small intestinal nematodes from captive gazelles in Spain. J Helminthol. 2001; 75:363-365.
- Pugh DG, ed. Sheep and Goat Medicine. 1st ed. College of Veterinary Medicine, Auburn University, Alabama, 2002.
- Schnieder T. Helminthosen der Wiederkauer. In: Schnieder T, eds. Veterinarmedizinische Parasitologie, Vollstanding Überarbeitete und Erweiterte Auflage, Parey, Germany. 2006:166-234.
- 19. Scott I, Hodgkinson SM, Lawton DEB, et al. Infection of sheep with adult and larval Ostertagia cimcuncincta. Intl J Parasitol. 1998; 28:1393-1401.
- 20. Scott I, Mckellar QA. The effects of excretions/secretions of Ostertagia cimcuncincta on ovine abomasal tissue in vitro. Intl J Parasitol. 1998; 28:451-460.
- 21. Shaw DJ, Dobson AP. Patterns of macro parasite abundance and aggregation in wildlife populations: a quantitive review. Parasitology, 1995; 111: I11-133.
- 22. Soulsby EJL, ed. "Helminths, Arthropods & Protozoa of Domesticated Animals," 7th Edition, Bailliere Tindall, London, 1986.
- 23. Stear M, Bairden K, Bishop SC, et al. The processes influencing the distribution of parasitic nematodes among naturally infected lambs. Parasitology. 1998, 117:165-171.
- 24. Stear M, Piedrafita D, Sloan S, et al. Teladorsagia circumcincta. WikiJournal of Science, 2019; 2(1):4.
- 25. Sutherland I, Scott I, eds. Gastrointestinal Nematodes of Sheep and Cattle, Biology and Control. Wiley-Blackwell, United Kingdom; 2010.
- 26. Taylor MA, Coop RL, Wall RL. Veterinary parasitology, 3rd edn. Blackwell, Oxford, 2007.
- Urquhart GM, Armour J, Duncan JL, Dunn AM, Jenning, FW, eds. Veterinary Parasitology. 2nd ed. Blackwell Science Ltd., Oxford; 1996:224-234.
- Zachary J. Pathologic basis of veterinary disease expert consult. In: Gelberg HB, ed. Alimentary System and the Peritoneum, Omentum, Mesentery, and Peritoneal Cavity. 6th ed. Mosby; 2017:391-401.